Bronchoalveolar Lavage Proteomics in Patients with Suspected Lung Cancer

Carvalho et. al. 2017

Presented by: Ilana Kelsey Jason Lin Mike MacArthur Molly McNamara

Outline

- Lung cancer and recent developments in non-invasive diagnosis
- Use of omics to identify cancerous from non-cancerous lesions
- Present study: attempt to extend methods to prospective study
- Findings and discussion about future directions
- Potential impact and caveats

Some lung cancer stats



Ferlay et. al. 2015

Diagnosis of lung cancer

- Advances in non-surgical diagnosis: transbronchial lung biopsy (TBLB), computed tomography (CT)
- Advances in earlier diagnosis
 - Chest radiography or low-dose CT scan
- Limitations
 - TBLB takes several samples, could be less representative
 - TBLB may miss lesions in peripheral areas of the lung
- Combining these traditional bronchoscopic diagnostics with BAL increases diagnostic rate



http://rizwan-nurani.com/lungcancer-diagnosis-.html

What is bronchoalveolar lavage (BAL)?





http://www.mdguidelines.com /bronchoscopy

https://www.olympus-europa.com/

Why BAL?

- Less invasive than surgery
- Can get a large amount of information from the samples
- Genomic and proteomic analysis have been established
- Highly specific although less highly sensitive



Oumeraci et. al. 2011

Objectives of this study

- Use BAL to analyze potential cases of lung cancer *prospectively*
- Identify further areas to explore for increased diagnosis rate

Study participants



Figure 1

Study participants







Study participants



Liquid Chromatography Tandem Mass Spectrometry



database

spectrometer (MS2 scan)

Liquid Chromatography Tandem Mass Spectrometry

- Protein isolated from bronchoalveolar lavage fluid
- UPLC Column separation
- Tandem MS
 - Data dependent acquisition
 - Top 6 hits per scan ionized
 - 60 second dynamic exclusion
- Database matching MaxQuant and VEMS
- Peptide/protein quantification
 - Spectral counts
 - Integration-based intensity values

Figure 2: PCA Analysis





Figure 2: PCA Analysis



Figure 2: PCA Analysis



Figure S7: Hierarchical Clustering



Figure 3: Boxplot of iBAQ expression values for the nine most significant regulated proteins

<u>Method</u>: LC-MS, Label-free quantification (iBAQ)

- Two search engines: MaxQuant and VEMS
- Filtering of proteins
 - 1. Benjamini & Hochberg method to decrease false positives (p<0.05)
 - 2. > two-fold regulation
 - Same direction of regulation in both MaxQuant and VEMS

<u>Takeaways:</u>

- 133 significantly regulated proteins identified
- Top 9 shown here (all are upregulated in lung cancer)
- Potential markers for lung cancer



Figure 4: GO enrichment analysis of all identified proteins

В

Α Enriched cellular components for all identifications 500 Number of protein identifications 400 300 200 00

Enriched cellular components for 133 significant regulated proteins 60 Number of protein identifications 50 40 30 20 10 0

<u>Method</u>: Gene Ontology analysis and binning of the significantly regulated proteins in their respective cellular components

<u>Takeaway:</u> Most of the proteins were from the extracellular vesicular exosome, and this enrichment was compared to the relative enrichment estimates, p values and number of identified proteins.

Figure S19

• Further characterization of the GO proteins



Figure 5: Overview of potential lung cancer biomarkers up- or down- regulated

<u>Method</u>: Heat map that compared biomarkers identified in this study to those in literature

Takeaways:

- Lack of consistency between tissue, BAL, BAL cells, and plasma
- Metabolic pathways most significant among differentially regulated genes
 - Possibility of fluorescently tagging metabolic enzyme inhibitors and using them for diagnostic purposes
- Other important KEGG pathways: spliceosome, focal adhesion, Epstein-Barr Virus infection



Figure 6



<u>Takeaways:</u> Cancer and Lung cancer have certain proteomes and certain significantly regulated proteins identified from lung cancer tissue from this study have been shown to be novel.

Strengths/Weaknesses

- Strengths:
- Heterogenous clinical population
- Moderate discriminatory ability for cases vs controls
- Potential to augment current diagnostic tools

- Weaknesses:
- Validating against difficult clinical diagnosis
- No classification models used
- Not diagnostic alone

Conclusions

- Demonstrate potential for biomarker diagnosis of lung cancer
 - Although they fall short of true diagnostic
- Some overlap is seen between their results and past studies
 - Mostly in upregulated protein
- Identification of specific cell compartments with differences between tumor and normal point to ways to refine the diagnosis rate

References

- Ferlay, J. *et al.* Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International journal of cancer* **136**, E359–386, doi: 10.1002/ijc.29210 (2015).
- Oumeraci, T. *et al.* Bronchoalveolar lavage fluid of lung cancer patients: mapping the uncharted waters using proteomics technology. *Lung Cancer* **72**, 136–138, doi: 10.1016/j.lungcan.2011.01.015 (2011).
- Almatroodi, S. A., McDonald, C. F., Collins, A. L., Darby, I. A. & Pouniotis, D. S. Quantitative proteomics of bronchoalveolar lavage fluid in lung adenocarcinoma. *Cancer genomics & proteomics* 12, 39–48 (2015).
- Carvalho, A. S., et al. (2017). "Bronchoalveolar Lavage Proteomics in Patients with Suspected Lung Cancer." <u>Scientific Reports</u> 7: 42190.