LS 50: Integrated Science

Course overview

LS50 is a two-semester, double course that introduces the natural sciences as an integrated whole. Its goal is to teach students how to solve scientific problems by drawing methods and concepts from biology, chemistry, physics, and mathematics. The course uses examples from biology as an integrating theme, principles from physics and mathematics to reduce complex problems to simpler forms, and computer simulation to allow students to develop their intuition about the behavior of the dynamical systems that control the physical and biological universe. The course includes bootcamps to introduce students to biological experiments and the computer language, MATLAB. Each semester will include a project lab, in which students will work in small teams to do original research on unsolved biological problems.

Natural science is a single intellectual enquiry into the universe of objects that surround us. Its components are linked by a common method: inducing hypotheses from a mixture of data and intuition, deducing predictions, and testing them by experiment and observation. The sciences depend on mathematics, from the simple act of counting to sophisticated methods required for computational chemistry and theoretical physics. The Integrated Science curriculum will introduce motivated freshmen to the concepts and methods needed to attack the life sciences in the 21st century. For both semesters, students will take the equivalent of two courses, meeting for formal instruction every day, performing hands-on, original research, and using modern computer methods to simulate scenarios and analyze data.

Darwin and Wallace's theory of evolution revealed that the living things have a purpose: their structure, function, and behavior are integrated to leave as many progeny as possible. For much of the 20th century, this difference, and the astonishing diversity of form and function, tended to separate biology from the other natural sciences: biology's complexity made it unappealing to many mathematicians, physicists, and chemists, and the "assume a spherical cow" flavor of theorists' simplifying assumptions made biologists skeptical about how useful theory was for understanding biology. Two advances have pushed biologists towards theorists and computer scientists: the need to test our understanding of biological processes by making explicit, mathematical models and the need to convert large datasets into information and, ultimately, knowledge.

We will teach students that the answer to "How will you solve this problem?" is "By any means necessary!" Our goal is to teach them how to find interesting problems, the means to solve them, and above all, the knowledge and courage to invent the new methods that make previously insoluble problems soluble. Coupling concepts and methods to problems that excite students and making them use these tools in their own research will embed the concepts in their working memory.

We will teach through iterated cycles of experiment and analysis, making use of experimental computation to simulate a system of interacting entities and explore the effect of parameter variation on the system's properties. Our goal is to complement the formal derivation of theorems, show the productive interplay between theory, simulation, and experiment, and show that computer systems and programs, like biological objects, have purposes. Mastering a restricted syntax to write algorithms will help students think about how biological systems use the restricted syntax of chemistry and genetics to accomplish tasks. Concepts like modularity, exploration with selection, error detection and correction, and recycling previous inventions are

important in the function and evolution of both organisms and code. Six faculty will teach the course, working as three pairs of one life scientist and one physical scientist.

Students will use their knowledge to conduct original scientific research. To ensure that all the students start with abilities in laboratory experiments and computer simulations, we will have two, two week-long boot camps, one in computation and one in experimentation. After the boot camps, students will perform original research in project laboratories. The project labs will be based on the research of and run by the Bauer Fellows, independent scientists who spend five years at Harvard after their PhDs and run small research groups.

Class format

Lectures each weekday (MTuWThF) from 10-11:30 in Northwest Labs B108 (first basement level), 52 Oxford St.

Weekly discussion sections with teaching fellows, at locations and times to be determined based on student availability.

Weekly problem sets, to be submitted as indicated on each assignment.

Two in-class tests during semester, each covering the preceding month's worth of lecture material.

Cumulative final exams, with particular emphasis on material covered in the last month of the semester.

Two, two week bootcamps: one in programming in Matlab, one in experimental biology Post-bootcamp, research-based laboratory: students work on original research projects in small groups, with open access to a sophisticated modern research laboratory. Students are expected to spend 6 hr/week working on their projects. Labs will meet Monday and Friday from 1-4 PM, beginning Monday, September 14th, in the teaching labs on the first basement level of Northwest Labs (52 Oxford St.). Bootcamps will include weekend sessions.

Faculty

An up-to-date list of contact information and office hours can be found on the course website at: https://canvas.harvard.edu/courses/3013/pages/course-staff-contact-information

Faculty

Ben de Bivort OEB
Michael Desai OEB
Cassandra Extavour OEB
Erel Levine Physics

Andrew Murray MCB, Course Head

Mary Wahl MCB

The year-long course will be divided into thirds, each team-taught by a pair of faculty, one from the physical and one from the life sciences. Murray will attend all the lectures to ensure continuity. We expect that the intense nature of the course will produce strong interactions both amongst the students and between them and the faculty.

Teaching Fellows and Assistants

| Mariela Petkova | Biophysics | Early fall |
|------------------|-----------------|--------------|
| Emma Nagy | MCB | Early fall |
| Cara Weisman | Biophysics | Late fall |
| Shu Wang | Biophysics | Late fall |
| Jim Valcourt | Systems Biology | Early spring |
| Parris Humphrey | OEB | Early spring |
| Zachary Werkoven | MCB | Late spring |
| Emily Hager | MCB | Late spring |

Laboratory Heads

| John Calarco | Bauer Fellow | Fall |
|------------------|--------------|--------|
| Lauren O'Connell | Bauer Fellow | Spring |

Laboratory Teaching Fellows and Assistants

| Xico Gracida | Systems Biology | Fall |
|--------------|-----------------|------|
| Adam Norris | Systems Biology | Fall |

Lecture content

The list that follows was developed during a single semester Graduate Seminar in Undergraduate Education, in which the six faculty who will teach the course and seven graduate students, most of whom will serve as teaching fellows for the course, participated. Since the course will meet every day for two semesters, it will have roughly 125 lectures. As a result the description below is extremely telegraphic, including field-specific words, phrases, and acronyms. To provide more detail, we include detailed descriptions of the materials from three lectures: the opening lecture, the first lecture on probability and inference, and a lecture from roughly a quarter of the way into the course on energy. All three introduce areas, rather than going into areas in depth, since we thought it would be useful for the committee to see material that non-scientists could usefully opine on. For all three, we have provided a one page description of the key material the lecture covers (a roadmap) and the slides. This material is assembled in a single PDF file.

| Lecture title | Lecture content |
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| Introduction & course overview | What do cells & organisms do to survive & reproduce? |
| Biology as computation | The Central Dogma, coding & decoding, sensation & response |
| Inferences, Rules, Errors | The scientific method, measurement, measurement & statistical errors |
| Probability distributions and Bayesian analysis | Probability distributions, frequentist vs. Bayesian approaches, the Central Limit Theorem |
| Equilibrium constants and energy diagrams/equilibrium vs. steady-state | Equilibria, on & off rate constants, kinetics, activation energy |
| Lac operon I: Introduction | History, genetics & chemistry, non-genetic individuality via bistability |
| Lac operon II: Paradigm for gene regulation | Search times, introduction to 1-D diffusion, modern analysis |
| Quantization, elements, atomic orbitals | Atomic models: Bohr, Schrodinger |
| Periodic trends, molecular orbitals | Electronegativity, hybridization, bonds |
| Organic molecules and chirality | Structure of organic molecules, chirality, complexity |
| Protein structure, intermolecular forces | Hydrogen and van der Waals bonding, protein structure |
| Macromolecules and membranes | Sugars, lipids, nucleic acids, membranes, membrane properties |
| Introduction to ordinary differential equations | What is a differential equation, solutions to simple separable equation how to code numerical solutions |
| Dynamical systems I: fixed points and phase plane analysis | 1-D dynamical systems: fixed points, phase plane analysis |
| Introduction to the cell cycle | Introduction, coordinating growth with division, yeast & embryos |

| Genetic and physiological analysis of the cell cycle | Embryonic oscillators vs. yeast dependent event chains, conserve regulators |
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| Molecular mechanism of the cell cycle | Cell cycle engine mechanism & regulation, cell cycle checkpoints |
| DNA replication as an example of a cell cycle-controlled process | Biology of DNA replication and repair, kinetic proofreading |
| Cell cycle regulation by external events | Cell cycle regulation by environmental and developmental inputs |
| Cell cycle regulation of events in time and space | Controlling events in time and space, polarization, mitosis |
| Electrostatics and Debye screening | Managing backbone electrostatic repulsion during DNA compaction: introduction to electrostatics, Debye screening, nucleand chromosomal organization |
| Math primer: multivariate, Taylor series | Functions of more than one variable, partial derivatives, Taylor series |
| Many particle systems, approach to equilibrium | Definitions of system, environment, and entropy; mechanical, chemical, thermal |
| Equilibrium and entropy | Definition of entropy in terms of multiplicity, intensive vs. extensi variables, equations of state |
| Work and heat, energy, free energy | Work and heat; first law of thermodynamics; adiabatic and isothermal processes |
| Energy budget of the cell | Magnitude of overall ATP flux and distribution between processes efficiency |
| Boltzmann statistics | Finding extrema under constraints; Lagrange multipliers; the Boltzmann distribution; application to air pressure at altitude |
| Two-state systems | Ligand-receptor binding; Schottky; paramagnetism |
| Ideal gas, ideal solid, equipartition theorem | Average energy of molecule in a gas; equipartition theorem |
| Law of mass action | Reaction rate does not often reflect apparent stoichiometry: relationship between reaction rate and mechanism; rarity of termolecular processes; Arrhenius equation |
| Specificity: ligand, receptors, homologous recombination | Relative binding energy of correct and incorrect pairings; limitations on accuracy in translation; revisiting lac repressor movement |
| Biochemical approach to cooperativity | Derivation of Adair equation and Hill curve from expressions for chemical equilibria; MWC and KNF models and their biological motivations |
| Statistical mechanics approach to cooperativity | Gibbs distribution and grand partition function; application to derive equivalent expressions for cooperativity. Applied to nuclei acid denaturation. |

| | Relationship between potentials and conservative forces; Fick's 1 |
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| Introduction to diffusion | and 2nd laws; Green's function for Brownian motion |
| Gaussian integrals, convolution, and applications to diffusion | Gaussian integrals and calculation of RMS distance traveled, convolution, applications to FRAP and other diffusion problems |
| Transport in the cell: diffusion, anomalous diffusion, and active transport | Detection of crowding/tethering and measurement of diffusion coefficient by FRAP, "speed" of diffusion, need for active transpor |
| Diffusion to detection (Berg and Purcell) | How many transporters are necessary? Steady-state solution for diffusion of particles towards absorbing regions on a sphere |
| Lac operon III: Application of rate equations and diffusion; revisiting classic system with modern techniques | Behavior of the lac operon explained via statistical mechanics, detailed analysis of diffusion and capture of the Lac repressor by the Lac operator |
| Molecular implementation of regulation: promoters, activators, repressors, chromatin | How transcription factors and cis-regulatory elements work (bacteria vs. eukaryotes), chromatin modification |
| Synthetic biological circuits | Design and implementation of synthetic circuits, e.g., genetic switches, repressilator, Collins toggle switch |
| Development I: discerning differences between cells | Methods for detecting differences in gene expression between cel epigenesis vs. predeterminism, overview of metazoan embryolog |
| Worms fate map (vulva as example) | Lineage, fate decisions, and noise in C. elegans vulval developmen |
| Chemical and phase equilibria | Clausius-Clapeyron, surface tension, mixing of hydrophobic molecules and water |
| Phase transitions | Phase diagrams, metastability, critical point, nucleation of solids |
| Solvation, solutions, and mixtures | Solvation, solutions, osmotic pressures, water and hydrophobicity |
| Electrostatics and the Nernst equation/intra-molecular interactions | The superposition principle; ionic bonds, hydrogen bonds, Londo dispersion forces can all be explained by Coulombic interactions; dipoles and dielectric effect; Bjerrum length calculation |
| Structure I | Modularity, primary through tertiary structure, turns, domains |
| Structure II | Self-assembly, Anfinsen, Levinthal, chaperones |
| Structure III | Driving forces of folding, exploration with selection (MCMC) |
| Introduction to optics | Reflection and refraction, Snell's law, lenses and microscopy; diffraction, Bragg's law |
| Fourier transforms | Definition of Fourier transform, examples, PSD for shot noise and relation to Poisson statistics, flicker and Brownian noise |
| Diffraction | Structure factor and relationship to electron density, diffraction grating demo, methods for phase determination |
| X-ray crystallography and EM | Interpretation of photo 51, introduction to EM |
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| Membranes, permeability, and pumps | Lipid structure, surface tension and surfactants, permeability, partition coefficients, pumps and channels |
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| Energy harvesting in photosynthesis | Introduction to photosynthesis with emphasis on light-dependent steps; production of proton gradient and ATP synthesis by relievi confinement. |
| Chemiosmotic hypothesis | Mitochondria, oxidative phosphorylation, generation of heat from brown fat, decoupling weight loss aids |
| Carbon fixation and consumption | Remaining steps of photosynthesis and glycolysis discussed |
| Enzymes I | Measurement, catalysis, local concentration |
| Enzymes II | Rate equations, separation of timescales, Michaelis-Menten kineti |
| Enzymes III | Orientation, arrow pushing, covalent intermediates |
| Two-component signaling | Introduction to signaling, crosstalk avoidance, arrow-pushing mechanism of HK-RR phosphotransfer |
| Linear algebra I | Vectors, scalars, vector space, dimensionality, component representation, matrices |
| Linear algebra II | Matrix multiplication, identity operator, and inverse |
| Linear algebra III | Change of basis, orthogonality |
| Linear algebra IV | Eigenvalues and eigenvectors |
| Principal Component Analysis (PCA) | Changing basis in a meaningful way, dimensionality reduction, application to genetic variation in Europeans |
| PCA application I: facial recognition | PCA applied to facial recognition |
| PCA application II: microbial communities | PCA applied to microbial communities, discussion of clustering methods |
| Newton's laws of motion | F=ma, second order differential equations, simple examples |
| Simple harmonic motion | Harmonic equation, solutions to this equation, ball on a spring. |
| Soft matter springs | Elasticity and entropic string |
| Conservation laws I | Energy and momentum |
| Conservation laws II | Example problems |
| Damped harmonic motion | Damped harmonic equation, overdamped and underdamped solutions |
| Driven harmonic motion | Driven harmonic equation, solution, resonance |
| Systems of harmonic equations | Normal modes of a system of harmonic equations |
| Examples of systems of harmonic equations | Coupled balls on a string, coupled pendulums, applications to modes of molecular oscillations |
| Motors and polarity | Directional motion of motors along polymers, dynein, sarcomere cycle |

| Thermodynamic cycles and | Carnot cycle, calculation of efficiency of ATP synthase motor, |
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| molecular motor efficiency | Brownian ratchets and directed motion |
| Cytoskeleton I | Basics, polymerization, critical concentration, spindle assembly |
| Cytoskeleton II | Non-equilibrium polymers |
| Segregation | Pros and cons of putting different activities in different compartments |
| Endosymbiont theory | Cyanobacterial/alpha-proteobacterial origin of the chloroplast/mitochondrion, gradual gene transfer, non-Mendeliai inheritance |
| Import and export | Nuclear transport, protein secretion |
| Symmetry and symmetry breaking | Symmetry as invariance under transformation, CP violation and CPT symmetry, lateral inhibition and bud site determination |
| Development II | A-P axis patterning in Drosophila embryo, maternal effect/gap/segment polarity genes, morphogens, useful transgeni techniques |
| Dynamical systems II | Bifurcations; examples possibly including spruce budworm, firefly flashing, homochirality |
| Dynamical systems III | Two-dimensional dynamical systems, classifying fixed points, spin nodes |
| Dynamical systems IV | Limit cycles, Lotka-Volterra, Poincare-Bendixson |
| Introduction to neuroscience | Membrane potentials, chemical synapses, gap junctions, calcium compartments |
| Hodgkin and Huxley | Hodgkin-Huxley model in neuroscience, solutions by simulations. |
| Optogenetics, behavioral feedback loops | Using light to control neural activity and probe the function of nervous systems |
| Locomotion | Pattern recognition, inhibitory-excitatory alternation, proprioception, efference copy |
| Stochastic processes: from random walks to Markov chains | Formalizing random walks as a Markov chain, basic properties of Markov chains |
| Stochastic differential equations | Langevin equation for a stochastic process, interpretations, simulations |
| Biological examples of stochasticity | Mutation, excitability, plasmid partition |
| Master equation, birth-death processes | Using matrices to model the temporal evolution of complex system |
| Noise in molecular biology | Stochasticity of gene expression, neuronal activity |
| Graph theory | Graphs as formal devices to represent connections within comple systems. Examples from control of neuronal activity and gene expression. |
| Network theory | Analysis of the structures and properties of large networks. |
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| Worms II | Connectome, EM reconstruction, network properties |
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| Metabolic networks | Metabolism as a dynamical system, flux balance analysis |
| Social interactions | Origins of complex behaviors (e.g. flocking) from simple rules |
| Classical genetics | Genes, alleles, sex, meiosis, mapping, screens |
| Molecular genetics I | Mutant selection, epistasis, chromosome elements, manipulation |
| Molecular genetics II | Diploids, transgenesis, CRISPR |
| Genomics | Sequencing, assembly, homology, BLAST |
| Evolutionary relationships | Phylogenetics, orthology, paralogy |
| Origin of life | Origin of life, RNA world |
| History of terrestrial biology and chemistry | History of life on earth, mass extinctions, major transitions in evolution |
| Diversity of life | Diversity of life, strange phyla |
| Population genetics I | Mutations and the origin and maintenance of diversity, selection, drift |
| Population genetics II | Sex: meiosis, sexual selection, Muller's ratchet |
| Population genetics III | Speciation and adaptive radiations |
| Population genetics IV | Detecting selection: MK, extended haplotype tests, bulk segregant/QTL |
| Evolutionary thinking | Drift, modification, recycling with modification, modularity |
| Molecular evolution of proteins | Evolution of protein function, extracting information from covariation |
| Evolution of development | Regulatory vs. coding region evolution, interpretation of heterologous expression experiments in Drosophilids, QTL applie to sticklebacks, duplication and divergence in coding regions |
| Diversifying mechanisms | Diversity inducing mech., contingency loci, switches, non genet individuality |

Assessment

Assessment will be based on five criteria:

In-class tests

We will administer two in-class tests, each covering the material presented in the preceding month. The question format and content will be similar to the weekly problem sets.

Final exams

Since this is a double class, we will administer two, three-hour finals in each semester. For each the goal will be that well-prepared students should be able to complete the final in two hours and questions will focus on solving problems (mathematical analysis of problems, data interpretation, and experimental design), rather then regurgitation. The final exam will be cumulative but

special emphasis will be placed on topics from the last month of the semester (since no in-class test will cover that material).

Problem sets

Weekly problem sets will review the topics covered in lectures and discussion sections. We highly encourage students to collaborate to solve the problems, which may include a comparison of scratch work, methods used, and numerical results. All submitted work should be written independently and reflect the students' own understanding. For example, a student should not submit work which they would be unable to explain or reproduce on their own.

This policy reflects our intent in assigning the problem sets: to help students master the material under low-stress circumstances so they will not be blindsided by higher-stakes tests. While one should be wary of over-reliance, classmates are an excellent resource for help since they keep similar hours, live nearby, and remember their own learning process well enough to explain concepts appropriately. On the flip side, mentoring colleagues whenever possible is not merely kind and forward-thinking, but also helps crystallize your understanding and practice the mentorship and presentation skills needed for work in the scientific community.

Laboratory research

Students will be graded on the research projects they undertake, focusing on their ability to design and interpret experiments and their commitment to and engagement with their research more than on the quality of the results they produce (subject as these are to the whims of the Gods). Approximately half of the grade from laboratory research will reflect weekly participation in lab, with the remainder attributed to a final lab report.

Lecture participation

Students will be graded on their participation in class, which we expect to be highly interactive, weekly conferences, and the discussions associated with laboratory research.

These criteria will be accorded the following weights

| In-class tests | 20% |
|-----------------------|-----|
| Final exams | 30% |
| Problem sets | 20% |
| Laboratory research | 20% |
| Lecture participation | 10% |